

PEROXIDASE ACTIVITY AND ISOENZYME PATTERN IN TOBACCO PLANTS EXPOSED TO SILVER NANOPARTICLES AND SILVER NITRATE

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INTRODUCTION

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With a rapidly growing implementation of nanomaterials in various consumer products, comes an even greater environmental concern. Silver nanoparticles (AgNPs) are the most commonly used nanomaterial in consumer products due to their antibacterial and antifungal properties. However, being released to water or soil, AgNPs are likely to interact with plants, which are a vital part of the ecosystem, and thereby enter the food chain. Although AgNPs are known to induce toxicity in prokaryotic organisms, their effect on plants has not been fully elucidated.¹ In this study, tobacco (*Nicotiana* tabacum) plants were simultaneously exposed to the same concentrations (25, 50, 75, 100 and 500 μ M) of AgNPs and ionic silver (AgNO₃), in order to determine AgNPs potential phytotoxic effects on this economically important crop plant. In terms of oxidative stress, changes in activities of antioxidant enzymes, pyrogallol (PPX) and ascorbate peroxidase (APX), were determined in both roots and leaves and compared to non-exposed, control plants. Furthermore, changes in PPX and APX isoforms expression in treated plants were observed in regard with Ag form and its concentration and compared to control.

MATERIALS AND METHODS

RESULTS

PPX activity roduct min⁻¹ mg⁻¹

25

20

10

Κ

25 µM

Table 1. Concentration of Ag measured in roots and leaves of adult tobacco plants. The results represent the mean value of 6 replicates ± standard error. Values





marked with different letters represent significant difference ($p \le 0.05$) according to Duncan test. Value < 0.0001 µg g⁻¹ represents instrument quantification bound

| | (µg g ⁻¹ _{fresh weight}) | (µg g ⁻¹ _{fresh weight}) |
|--------------------------|---|---|
| Control | < 0.0001ª | < 0.0001ª |
| 25 μM AgNPs | 1247.4 ± 122.0 ^b | 12.1 ± 2.6 ^b |
| 50 μM AgNPs | 1395.2 ± 351.5 ^b | 13.9 ± 2.6^{b} |
| 75 μM AgNPs | 1712.1 ± 80.8 ^{bc} | 19.0 ± 1.0^{b} |
| 100 μM AgNPs | 1742.2 ± 192.8 ^{bc} | 36.3 ± 3.4° |
| 500 μM AgNPs | 2480.1 ± 141.9 ^d | 79.2 ± 5.2 ^d |
| 25 μM AgNO ₃ | 1121.5 ± 136.2 ^b | 18.2 ± 4.0^{b} |
| 50 μM AgNO ₃ | 1450.9 ± 436.2 ^b | 21.4 ± 7.3 ^b |
| 75 μM AgNO ₃ | 1741.7 ± 134.5 ^{bc} | 23.0 ± 4.1 ^b |
| 100 μM AgNO ₃ | 1747.4 ± 150.0 ^{bc} | 38.3 ± 5.4° |
| 500 μM AgNO ₃ | 2399.1 ± 310.6 ^{cd} | 82.2 ± 3.9 ^d |

AgNO3 AaNP

100 µM

500 µM

■AgNP ■AgNO3



Figure 2. Activity of APX in tobacco roots after exposure to 25, 50, 75, 100 and 500 μ M AgNPs and AgNO₃. The presented results show mean values of 6 replicates ± standard error. K control. Values marked with different letters represent significant difference ($p \le 0.05$) according to Duncan test.



75 µM

50 µM

significant difference ($p \le 0.05$) according to Duncan test.

Figure 1. Activity of PPX in tobacco roots after exposure to 25,

50, 75, 100 and 500 μ M AgNPs and AgNO₃. The presented

results show mean values of 6 replicates ± standard error. K -

control. Values marked with different letters represent



ANALYSES:





Figure 3. Activity of PPX in tobacco leaves after exposure to 25, 50, 75, 100 and 500 μ M AgNPs and AgNO₃. The presented results show mean values of 6 replicates ± standard error. K control. Values marked with different letters represent significant difference ($p \le 0.05$) according to Duncan test.



Figure 5. PPX isoforms of tobacco roots after conducted native PAGE. K - control; 1 - 25 μM, 2 - 50 μM , 3 - 75 μM, 4 - 100 μM and 5 - 500 μM AgNPs; 6 - 25 μM, 7 - 50 μM, 8 - 75 μM, 9 - 100 μ M and 10 - 500 μ M AgNO₃.





Figure 4. Activity of APX in tobacco leaves after exposure to 25, 50, 75, 100 and 500 μ M AgNPs and AgNO₃. The presented results show mean values of 6 replicates ± standard error. K control. Values marked with different letters represent significant difference ($p \le 0.05$) according to Duncan test.

K 1 2 3 4 5 6 7 8 9 10



Figure 6. PPX isoforms of tobacco leaves after conducted native PAGE. K - control; 1 - 25 μM, 2 - 50 μM , 3 - 75 μM, 4 -100 μM and 5 - 500 μM AgNPs; 6 - 25 μM, 7 - 50 μM, 8 - 75 μ M, 9 - 100 μ M and 10 - 500 μ M AgNO₃.





REFERENCES

- 1. Cvjetko et. al. (2018), Environ Sci Pollut Res Int 25:5590-5602.
- 2. Murashige and Skoog (1962), Physiologia Plantarum 15: 473-497.
- 3. Mittler and Zilinskas (1993), Analytical Biochemistry, 212:540-546.
- 4. Nakano and Asada (1981), Plant Cell Physiol 22: 867-880.
- 5. Chance and Maehly (1955), Methods in Enzymology 2:764-775.



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Figure 7. APX isoforms of tobacco roots after conducted native PAGE. K - control; 1 - 25 μM, 2 - 50 μM , 3 - 75 μM, 4 -100 μM and 5 - 500 μM AgNPs; 6 - 25 μM, 7 - 50 μM, 8 - 75 μ M, 9 - 100 μ M and 10 - 500 μ M AgNO₃.

Figure 8. APX isoforms of tobacco leaves after conducted native PAGE. K - control; 1 - 25 μ M, 2 - 50 μ M, 3 - 75 μ M, 4 -100 μM and 5 - 500 μM AgNPs; 6 - 25 μM, 7 - 50 μM, 8- 75 μM, 9 - 100 μ M and 10 - 500 μ M AgNO₃.

CONCLUSIONS

• After both types of treatments similar accumulation of Ag was obtained, which was significantly higher in roots than in leaves.

• Concentrations and the form of applied Ag seemed to be correlated with activities of both APX and PPX since significant differences were obtained between AgNP- and AgNO₃-treatments in both roots and leaves. • Peroxidase isoenzyme patterns exhibited that the Ag form and its concentration affect the peroxidase isoforms expression; namely, quantitative and qualitative changes were observed in PPX and APX isoforms in treated plants compared to control, but also in roots and leaves of plants exposed to different forms of Ag. • Obtained results suggest that AgNPs induce different and specific response in plant's antioxidant enzymes compared to $AgNO_3$.